

# Volatility in the effective size of a freshwater gastropod population

Robert T. Dillon Jr 

Department of Biology, College of Charleston, Charleston, SC, USA

## Correspondence

Robert T. Dillon, Jr, Freshwater Gastropods of North America Project, Charleston, SC, USA.  
Email: DillonR@fwgna.org

## Abstract

Despite the utility of gastropod models for the study of evolutionary processes of great generality and importance, their effective population size has rarely been estimated in the field. Here, we report allele frequency variance at three allozyme-encoding loci monitored over 7 years in a population of the invasive freshwater pulmonate snail *Physa acuta* (Draparnaud 1805), estimating effective population size with both single-sample and two-sample approaches. Estimated  $N_e$  declined from effectively infinite in 2009 to approximately 40–50 in 2012 and then rose back to infinity in 2015, corresponding to a striking fluctuation in the apparent census size of the population. Such volatility in  $N_e$  may reflect cryptic population subdivision.

## KEYWORDS

allozyme frequencies, effective population size, *Physa acuta*, population subdivision, pulmonate snails, simultaneous hermaphrodite

## 1 | INTRODUCTION

Since the introduction of the concept by Wright (1931), effective population size ( $N_e$ ) has been adopted as a parameter in scores of evolutionary models, adaptive and neutral alike (Crow, 2010). The concept has found important applications in animal breeding (Caballero, Santiago, & Toro, 1996) and in conservation biology (Nunney & Elam, 1994). Two categories of methods to estimate  $N_e$  from field data were developed in the early 1980s, a single-sample approach based on disequilibrium between alleles at unlinked loci and a two-sample approach based on variance in allelic frequencies between generations (Caballero, 1994). Several single-sample methods not relying on linkage disequilibrium have more recently been proposed.

Despite the importance of the concept, however, effective population size has rarely been estimated for any gastropod population in the field. Crow and Morton (1955) used variance in progeny number to estimate the  $N_e/n$  ratio in a laboratory culture of the freshwater pulmonate *Lymnaea (Pseudosuccinea) columella*. The earliest field estimates were those of Murray (1964) and Greenwood (1974), who

applied simple single-sample approaches to shell color polymorphism in an English population of the important land snail model *Cepaea nemoralis*. But another thirty years would elapse before estimates of effective population size were offered for other land snail populations, those of Arnaud and Laval (2004) using microsatellite markers and a two-sample method, and Ursenbacher, Alvarez, Armbruster, and Baur (2010) using a one-sample approach.

We are aware of two estimates of the effective size of marine gastropod populations, both inhabiting the European intertidal. Fernandez et al. (2005) followed variation at allozyme-encoding loci over 14 years in incipient species of *Littorina saxatilis*, estimating effective population size using a two-sample approach. Riquet, Le Cam, Fonteneau, and Viard (2016) analyzed microsatellite variation in an invasive population of *Crepidula fornicata* over 9 years, comparing both one-sample and two-sample estimates.

In the freshwater gastropods, Meunier, Hurtrez-Bousses, Durand, Rondelaud, and Renaud (2004) used both one-sample and two-sample analyses of microsatellite polymorphism to estimate the effective sizes of six French populations of the (predominantly self-fertilizing)

pulmonate *Lymnaea (Galba) truncatula*. Microsatellites and two-sample techniques were also used by Trouve, Degen, and Goudet (2005) on six populations of *L. truncatula* sampled from Switzerland. The literature contains two single-sample microsatellite studies on Chinese populations of viviparid snails—*Bellamya quadrata* (Gu, Zhang, et al., 2015) and *B. purificata* (Gu, Zhou, et al., 2015).

In recent years, the freshwater basommatophoran pulmonate snail *Physa acuta* has found widespread use as a model organism for a variety of evolutionary studies (Figure 1). Populations of *P. acuta* in both field and laboratory settings have played important roles in studies of mating behavior (Janicke, Vellnow, Lamy, Chapuis, & David, 2014; Janicke, Vellnow, Sarda, & David, 2013; Wethington & Dillon, 1996), sex allocation (Janicke & Chapuis, 2016; Wethington & Dillon, 1993), inbreeding depression (Jarne, Perdieu, Pernot, Delay, & David, 2000; Noel et al., 2016), reproductive isolation (Dillon, Robinson, & Wethington, 2007), gene flow (Bousset, Henry, Sourrouille, & Jarne, 2004; Van Leeuwen et al., 2013), speciation (Dillon, Wethington, & Lydeard, 2011), and ecophenotypic plasticity (Auld & Relyea, 2011; Dillon & Jacquemin, 2015; Gustafson, Kensinger, Bolek, & Luttbeg, 2014).

Native to North America, invasive populations of *P. acuta* have been introduced around the world and are now established on six continents, typically in rich, disturbed, and lentic environments (Albrecht, Kroll, Terrazas, & Wilke, 2009; Dillon, Wethington, Rhett, & Smith, 2002). The snail is simultaneously hermaphroditic and capable of self-fertilization (Dillon, McCullough, & Earnhardt, 2005), although outcrossing is preferred (Escobar et al., 2011; Wethington & Dillon, 1997). Generation time in the laboratory can be as short as 6 weeks (Wethington & Dillon, 1993), although wild populations typically complete only one or two generations per year, both effectively semelparous (Life cycles A or C of Dillon, 2000: 158).



**FIGURE 1** *Physa acuta* (9 mm shell length), courtesy D. Liebman

We originally sampled the population of *P. acuta* inhabiting the Quarterman Park “Duck Pond” in North Charleston, SC, as part of a 1991 population genetic survey of the Carolina Sea Islands (population “NPK” of Dillon & Wethington, 1995). The population demonstrated allozyme variation interpretable as the product of codominant alleles segregating in Mendelian fashion at three loci: isocitrate dehydrogenase (Isdh), 6-phosphogluconate dehydrogenase (6pgd), and esterase-3 (Est3). Mendelian inheritance at the (strong, slow) Est3 locus has been confirmed by Dillon and Wethington (1994).

The Duck Pond drains directly into the brackish Cooper River, effectively isolating the population of freshwater snails it contains by both land and sea. The nearest neighboring population of *P. acuta* is probably that inhabiting the upper, freshwater marshes of Filbin Creek, approximately 2 km north overland. Ducks and other waterfowl doubtless visit both habitats, providing some opportunity for genetic exchange, albeit infrequent. The effects of migration on  $N_e$  have been studied by Gilbert and Whitlock (2015).

## 2 | MATERIALS AND METHODS

The Quarterman Park Duck Pond (32.87822, -79.98077) was constructed from a marshy embayment of the Cooper River in the early 20th century. For most of its history, it was tidally influenced and slightly brackish, but recent drainage improvements have rendered it entirely fresh, fed by local runoff. Its area at present is approximately 1.0 hectare, and depth is no more than 2 m.

The pond is maintained by city personnel at irregular intervals and has been kept free of macrophytic vegetation in recent years. Water temperatures can rise above 35°C during summer months, depressing dissolved oxygen to low levels, despite city efforts at artificial aeration. Its population of *P. acuta* reaches maximum density on allochthonous leaves and debris floating at the eastern (windward) end of the pond, at the drain.

We visited the pond each spring from 2009 to 2015, beginning in March, examining debris at the eastern end to qualitatively assess snail densities. If the apparent census size was sufficient to yield several hundred snails with reasonable effort, an annual sample was taken. Otherwise, we postponed the sample and returned a few weeks later. Approximately 150–200 *P. acuta* were ultimately sampled every spring, with one exception. The exception was 2012, when the snail population never reached an abundance at which it could be sampled, from March to August.

Snails collected at each sampling year were returned to the laboratory and frozen individually in 80–160  $\mu$ l of tris tissue buffer for analysis of allozyme polymorphism. We used horizontal starch gel electrophoresis in a TEB8 buffer system to resolve variation at the Est3, Isdh, and 6pgd loci and an aminopropylmorpholine pH 6 buffer system for a second examination of Isdh and 6pgd. Details regarding our electrophoretic methods, including a description of our equipment and recipes for all stains and buffers employed, have been published by Dillon (1992) and Dillon and Wethington (1995).

Allele frequencies and tests of fit to Hardy–Weinberg expectation were calculated using GenePop version 4.5.1 (Raymond & Rousset, 1995; Rousset, 2008). Values of  $F_{IS}$  were computed using the method of Weir and Cockerham (1984), and exact  $p$ -values were by the Markov chain method.

NeEstimator v2.01 is freely available software designed to estimate effective population size using three single-sample methods and three two-sample (moment-based temporal) methods (Do et al., 2014). Among the single-sample methods, Gilbert and Whitlock (2015) reported that the linkage disequilibrium (LDNe) method of Waples and Do (2008) consistently returned the lowest root square mean error across the range of effective population numbers simulated, absent migration.

The three two-sample methods implemented by NeEstimator 2.01 employ the standard temporal method (ST) of Waples (1989), with different approaches to computing standardized allele frequency variance: the  $F_c$  of Nei and Tajima (1981), the  $F_k$  of Pollak (1983), and the  $F_s$  of Jorde and Ryman (2007). The simulations of Gilbert and Whitlock (2015) suggested that all three of these two-sample methods perform with equivalent efficiency. Thus, we elected to estimate the effective population size of the Quarterman Park *P. acuta* population using four approaches: LDNe, STFc, STFk, and STFs. Jackknife methods were used to calculate 95% confidence intervals (CI) for all  $N_e$  estimates.

### 3 | RESULTS

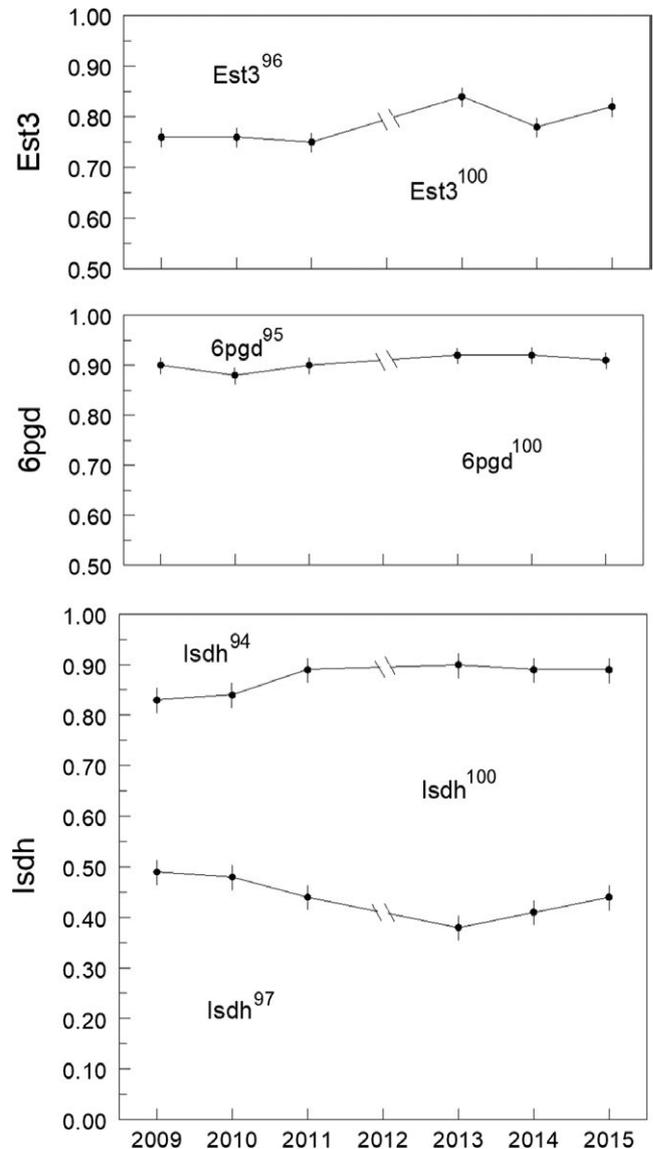
We resolved allozyme variation apparently encoded by two codominant alleles at the Est3 and 6pgd loci and three codominant alleles at Isdh. These alleles were named by the mobility of their allozyme bands relative to the standards set by Dillon and Wethington (1995) and plotted by their frequencies in Figure 2.

Sample sizes, values of  $F_{IS}$ , and values of  $p$  from goodness-of-fit tests to Hardy–Weinberg expectation are reported in Table 1. Over the entire data set of 3 loci  $\times$  6 years, 15 of the values of  $F_{IS}$  were positive, some strikingly so, and three were slightly negative. Four values of  $F_{IS}$  suggested significant heterozygote deficiencies (three at the Est3 locus and one at 6pgd), although not significant after Bonferroni correction.

The six single-sample estimates of effective population size based on linkage disequilibrium between the three loci are reported at the bottom of Table 1, with 95% CI. Table 2 shows the five two-sample estimates of effective population size based on allele frequency variance across pairs of consecutive samples. Each two-sample calculation was performed using  $F_c$ ,  $F_k$ , and  $F_s$  methods, yielding  $3 \times 5 = 15$   $N_e$  estimates, with confidence intervals.

### 4 | DISCUSSION

Tables 1 and 2 suggest that the effective size of the *P. acuta* population inhabiting the Quarterman Park Duck Pond was strikingly volatile, dipping from infinite in 2009 down to (remarkably consistent) values



**FIGURE 2** Allele frequencies at three allozyme-encoding loci in the Quarterman Park *Physa acuta* population, sampled over 7 years. Bars are 95% CI

of  $44.0 \pm 10.8$  by static estimate, or  $49.8 \pm 8.1$  by two-sample estimate, and then back up to infinity again. The dramatic fluctuation in  $N_e$  seemed to correspond to a fluctuation in apparent census size noticeable in the field, from (surely) thousands in the spring of 2009 to a very few in 2012, and then back up to thousands.

The spring of 2012 was exceptionally warm in North Charleston. The average temperature recorded by the National Weather Service over the month of March, 2012, was 65.3°F (18.5°C), the second highest mean March temperature in the 80-year record. It was our pond side observation that the *P. acuta* population at Quarterman Park never bloomed in the spring of 2012, which seemed to depress its size for the remainder of the year.

Both the effective size and the apparent census size of the *P. acuta* population in the Quarterman Duck Pond apparently returned to many thousands (at minimum) in just 2 years, perhaps four generations. So

the most obvious hypothesis to account for the depression in effective size observed between 2011 and 2013 would be a population bottleneck. But  $N_e$  is not expected to recover from such a striking bottleneck event until many generations have passed, absent migration (Caballero, 1994).

A less obvious hypothesis might be fluctuation in the selfing rate, such that the population of *P. acuta* in Quarterman Park shifted from outbreeding to inbreeding and then back again over the study interval. The only estimates of  $N_e$  remotely comparable to ours in the published literature are the works of Meunier et al. (2004) and Trouve et al. (2005) on European populations of the preferentially selfing *Lymnaea (Galba) truncatula*. The French populations studied by the former authors generally demonstrated  $N_e < 30$ , and the Swiss populations studied by the latter  $N_e < 10$ . But the selfing rates inferred for all *L. truncatula* populations in both studies, estimated from  $F_{IS}$ , consistently exceeded 80%. The heterozygosities we observed in our study population of *P. acuta* did not vary from expectation through our 7-year observation period.

We suggest that cryptic population subdivision may be the most likely hypothesis to account for the apparent volatility of  $N_e$  in our

7-year record. We sampled the Quarterman Park population of *P. acuta* at the east end of the pond for convenience. Snails were also observed elsewhere around the entire margin of the one-hectare pond, but not in densities sufficient to sample in the quantities required. Perhaps the striking dip in apparent population census size we observed in 2012 was localized on the east end, and its subsequent recovery was due to immigration from elsewhere within a subdivided population.

Some of the most influential studies of population subdivision published to date have been conducted using land snail models. Cain and Currey (1963) described small-scale variation in the frequencies of shell color morphs in the English land snail *Cepaea* as “area effects,” attributing the phenomenon to genetic drift. Among the earliest examples of parapatric speciation to be proposed was that of Murray and Clarke (1968), working with the localized clines in shell color polymorphisms demonstrated by the tropical land snail, *Partula*. Selander and Kaufman (1975) documented significant variance in the frequencies of allozyme-encoding genes in a population of the land snail *Helix (Cornu) aspersa* inhabiting two city blocks in Bryan, Texas. Some of this variance could be correlated with observable barriers to dispersal such as roads or walkways, but some could not.

The published literature also contains many reports of significant values of  $F_{ST}$  among subpopulations of freshwater snails. Most of these studies have been conducted where subpopulations are divided by identifiable barriers to dispersal, however, which is not the case in the Duck Pond at Quarterman Park. Jarne and Delay (1990) estimated values of  $F_{IS}$  and  $F_{ST}$  within and among several subpopulations of *Lymnaea peregra* (“*Radix balthica*”) sampled from Lake Geneva, reporting large values of the former but rather small values of the latter. Dybdahl and Lively (1996) reported significant values of  $F_{ST}$  between subpopulations of *Potamopyrgus antipodarum* sampled within several New Zealand lakes.

Like *Lymnaea truncatula*, the freshwater pulmonate snail *Bulinus truncatus* is a preferential self-fertilizer, Viard, Justy, and Jarne (1997) estimating selfing rates across 38 West African populations from 80% to 100%. The authors noted surprising variation in the levels of microsatellite polymorphism demonstrated by these populations, however, some showing no polymorphism at any of the four loci examined, others averaging over 10 alleles per locus. Viard and colleagues suggested some unseen variation in population sizes as a possible explanation for this phenomenon.

Puurtinen, Knott, Suonpaa, Van Ooik, and Kaitala (2004) reported significant positive correlations between the microsatellite polymorphism demonstrated by eight Finnish populations of the preferentially

**TABLE 1** Sample sizes, values of  $F_{IS}$ , and values of  $p$  from goodness-of-fit tests to Hardy–Weinberg expectation at three allozyme-encoding loci analyzed for the Quarterman Park *Physa acuta* population 2009–2015. The bottom two rows report single-sample estimates of effective population size. Inf., effectively infinite

	2009	2010	2011	2013	2014	2015
Est3						
N	217	217	217	184	217	186
$F_{IS}$	0.054	0.149	−0.002	0.184	0.207	0.060
$p$	.461	.034	1.00	.022	.005	.465
6pgd						
N	217	217	216	186	217	186
$F_{IS}$	−0.009	0.157	−0.057	0.053	−0.023	0.088
$p$	1.00	.029	.700	.366	1.00	.204
Isdh						
N	210	215	215	186	217	186
$F_{IS}$	0.032	0.069	0.114	0.037	0.090	0.107
$p$	.631	.568	.138	.673	.086	.079
LDNe	Inf.	Inf.	125.8	44.0	190.8	9,752
95% CI			34.6	10.8	2.0	51.0

**TABLE 2** Two-sample estimates of effective population size calculated for the Quarterman Park *Physa acuta* population 2009–2015, using three different approaches to estimate standardized allele frequency variance. Inf., effectively infinite

	2009–2010	2010–2011	2011–2013	2013–2014	2014–2015
STFc	Inf.	131.5	49.8	255.8	Inf.
95% CI		9.2	8.1	7.3	
STFs	Inf.	113.6	40.6	205.4	15,361
95% CI		34.8	17.3	35.9	453
STFk	Inf.	104.7	56.5	354.2	Inf.
95% CI		8.0	9.0	8.1	

outcrossing freshwater pulmonate, *Lymnaea stagnalis*, and several measures of fitness, including maturation age and fecundity. The authors suggested that their measures of genetic variation might indirectly estimate effective population size, with lower values of  $N_e$  promoting the random fixation of deleterious alleles. But Puurtinen and colleagues could not demonstrate a correlation between either genetic variability or population fitness and the current densities of the snail populations they sampled.

It should be cautioned that the number of genetic markers employed for the present study was small. The effectively infinite population sizes we estimated 2009–2010 and 2015 might result from fluctuating sampling variance. But our results, suggesting as they do striking volatility in the effective population size of a common and widespread pulmonate snail, offer a potential resolution to quandaries such as those reported by Viard, Puurtinen, and their colleagues.

## CONFLICT OF INTEREST

None declared.

## ORCID

Robert T. Dillon  <http://orcid.org/0000-0002-4224-536X>

## REFERENCES

- Albrecht, C., Kroll, O., Terrazas, E., & Wilke, T. (2009). Invasion of ancient Lake Titicaca by the globally invasive *Physa acuta* (Gastropoda: Pulmonata: Hygrophila). *Biological Invasions*, *11*, 1821–1826. <https://doi.org/10.1007/s10530-008-9360-9>
- Arnaud, J.-F., & Laval, G. (2004). Stability of genetic structure and effective population size inferred from temporal changes of microsatellite DNA polymorphisms in the land snail *Helix aspersa* (Gastropoda: Helicidae). *Biological Journal of the Linnean Society*, *82*, 89–102. <https://doi.org/10.1111/j.1095-8312.2004.00320.x>
- Auld, J., & Relyea, R. (2011). Adaptive plasticity in predator-induced defenses in a common freshwater snail: Altered selection and mode of predation due to prey phenotype. *Evolutionary Ecology*, *25*, 189–202. <https://doi.org/10.1007/s10682-010-9394-1>
- Bousset, L., Henry, P.-Y., Sourrouille, P., & Jarne, P. (2004). Population biology of the invasive freshwater snail *Physa acuta* approached through genetic markers, ecological characterization and demography. *Molecular Ecology*, *13*, 2023–2036. <https://doi.org/10.1111/j.1365-294X.2004.02200.x>
- Caballero, A. (1994). Developments in the prediction of effective population size. *Heredity*, *73*, 657–679. <https://doi.org/10.1038/hdy.1994.174>
- Caballero, A., Santiago, E., & Toro, M. (1996). Systems of mating to reduce inbreeding in selected populations. *Journal of Animal Science*, *62*, 431–442. <https://doi.org/10.1017/S1357729800014971>
- Cain, A., & Currey, J. (1963). Area effects in *Cepaea*. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, *246*, 1–81. <https://doi.org/10.1098/rstb.1963.0001>
- Crow, J. (2010). Wright and Fisher on inbreeding and random drift. *Genetics*, *184*, 609–611. <https://doi.org/10.1534/genetics.109.110023>
- Crow, J., & Morton, N. (1955). Measurement of gene frequency drift in small populations. *Evolution*, *9*, 202–214. <https://doi.org/10.1111/j.1558-5646.1955.tb01531.x>
- Dillon, R. T. Jr (1992). Electrophoresis IV, nuts and bolts. *World Aquaculture*, *23*(2), 48–51.
- Dillon, R. T. Jr (2000). *The ecology of freshwater molluscs*. Cambridge, UK: Cambridge University Press. <https://doi.org/10.1017/CBO9780511542008>
- Dillon, R. T. Jr, & Jacquemin, S. (2015). The heritability of shell morphometrics in the freshwater pulmonate gastropod *Physa*. *PLoS One*, *10*, e0121962. <https://doi.org/10.1371/journal.pone.0121962>
- Dillon, R. T. Jr, McCullough, T., & Earnhardt, C. (2005). Estimates of natural allosperm storage capacity and self-fertilization rate in the hermaphroditic freshwater pulmonate snail, *Physa acuta*. *Invertebrate Reproduction & Development*, *47*, 111–115. <https://doi.org/10.1080/07924259.2005.9652151>
- Dillon, R. T. Jr, Robinson, J., & Wethington, A. R. (2007). Empirical estimates of reproductive isolation among the freshwater pulmonate snails *Physa acuta*, *P. pomilia*, and *P. hendersoni*. *Malacologia*, *49*, 283–292. <https://doi.org/10.4002/0076-2997-49.2.283>
- Dillon, R. T. Jr, & Wethington, A. R. (1994). Inheritance at five loci in the freshwater snail, *Physa heterostropha*. *Biochemical Genetics*, *32*, 75–82. <https://doi.org/10.1007/BF00554416>
- Dillon, R. T. Jr, & Wethington, A. R. (1995). The biogeography of sea islands: Clues from the population genetics of the freshwater snail, *Physa heterostropha*. *Systematic Biology*, *44*, 400–408. <https://doi.org/10.1093/sysbio/44.3.400>
- Dillon, R. T. Jr, Wethington, A., & Lydeard, C. (2011). The evolution of reproductive isolation in a simultaneous hermaphrodite, the freshwater snail *Physa*. *BMC Evolutionary Biology*, *11*, 144. <https://doi.org/10.1186/1471-2148-11-144>
- Dillon, R. T. Jr, Wethington, A. R., Rhett, J., & Smith, T. (2002). Populations of the European freshwater pulmonate *Physa acuta* are not reproductively isolated from American *Physa heterostropha* or *Physa integra*. *Invertebrate Biology*, *121*, 226–234.
- Do, C., Waples, R., Peel, D., Macbeth, G., Tillett, B., & Ovenden, J. (2014). NeEstimator v2: Re-implementation of software for the estimation of contemporary effective population size ( $N_e$ ) from genetic data. *Molecular Ecology Resources*, *14*, 209–214. <https://doi.org/10.1111/1755-0998.12157>
- Dybdahl, M., & Lively, C. (1996). The geography of coevolution: Comparative population structures for a snail and its trematode parasite. *Evolution*, *50*, 2264–2275. <https://doi.org/10.1111/j.1558-5646.1996.tb03615.x>
- Escobar, J., Auld, J., Correa, A., Alonso, J., Bony, Y., Coutellec, M., ... David, P. (2011). Patterns of mating-system evolution in hermaphroditic animals: Correlations among selfing rate, inbreeding depression, and the timing of reproduction. *Evolution*, *65*, 1233–1253. <https://doi.org/10.1111/j.1558-5646.2011.01218.x>
- Fernandez, J., Galindo, J., Fernandez, B., Perez-Figueroa, A., Caballero, A., & Rolan-Alvarez, E. (2005). Genetic differentiation and estimation of effective population size and migration rates in two sympatric ecotypes of the marine snail *Littorina saxatilis*. *Journal of Heredity*, *96*, 460–464. <https://doi.org/10.1093/jhered/esi064>
- Gilbert, K., & Whitlock, M. (2015). Evaluating methods for estimating local effective population size with and without migration. *Evolution*, *69*, 2154–2166. <https://doi.org/10.1111/evo.12713>
- Greenwood, J. (1974). Effective population numbers in the snail *Cepaea nemoralis*. *Evolution*, *28*, 513–526. <https://doi.org/10.1111/j.1558-5646.1974.tb00785.x>
- Gu, Q.-H., Zhang, M., Zhou, C.-J., Zhu, G.-R., Dong, J., Gao, Y.-N., ... Chen, P. (2015). Analysis of genetic diversity and population structure of *Bellamya quadrata* from lakes of middle and lower Yangtze river. *Genetica*, *143*, 545–554. <https://doi.org/10.1007/s10709-015-9852-3>
- Gu, Q.-H., Zhou, C.-J., Cheng, Q.-Q., Li, X.-J., Zhu, G.-R., Zhang, M., ... Dong, J. (2015). The perplexing population genetic structure of *Bellamya purificata* (Gastropoda: Viviparidae): Low genetic differentiation despite low dispersal ability. *Journal of Molluscan Studies*, *81*, 466–475. <https://doi.org/10.1093/mollus/eyv017>

- Gustafson, K., Kensinger, B., Bolek, M., & Luttbeg, B. (2014). Distinct snail (*Physa*) morphotypes from different habitats converge in shell shape and size under common garden conditions. *Evolutionary Ecology Research*, 16, 77–89.
- Janicke, T., & Chapuis, E. (2016). Condition dependence of male and female reproductive success: Insights from a simultaneous hermaphrodite. *Ecology and Evolution*, 6, 830–841. <https://doi.org/10.1002/ece3.1916>
- Janicke, T., Vellnow, N., Lamy, T., Chapuis, E., & David, P. (2014). Inbreeding depression of mating behavior and its reproductive consequences in a freshwater snail. *Behavioral Ecology*, 25, 288–299. <https://doi.org/10.1093/beheco/art122>
- Janicke, T., Vellnow, N., Sarda, V., & David, P. (2013). Sex-specific inbreeding depression depends on the strength of male-male competition. *Evolution*, 67, 2861–2875.
- Jarne, P., & Delay, B. (1990). Inbreeding depression and self-fertilization in *Lymnaea peregra* (Gastropoda: Pulmonata). *Heredity*, 64, 169–175. <https://doi.org/10.1038/hdy.1990.21>
- Jarne, P., Perdieu, M.-A., Pernot, A.-F., Delay, B., & David, P. (2000). The influence of self-fertilization and grouping on fitness attributes in the freshwater snail *Physa acuta*: Population and individual inbreeding depression. *Journal of Evolutionary Biology*, 13, 645–655. <https://doi.org/10.1046/j.1420-9101.2000.00204.x>
- Jorde, P., & Ryman, N. (2007). Unbiased estimator for genetic drift and effective population size. *Genetics*, 177, 927–935. <https://doi.org/10.1534/genetics.107.075481>
- Meunier, C., Hurtrez-Bousses, S., Durand, P., Rondelaud, D., & Renaud, F. (2004). Small effective population sizes in a widespread selfing species, *Lymnaea truncatula* (Gastropoda: Pulmonata). *Molecular Ecology*, 13, 2535–2543. <https://doi.org/10.1111/j.1365-294X.2004.02242.x>
- Murray, J. (1964). Multiple mating and effective population size in *Cepaea nemoralis*. *Evolution*, 18, 283–291. <https://doi.org/10.1111/j.1558-5646.1964.tb01601.x>
- Murray, J., & Clarke, B. (1968). Partial reproductive isolation in the genus *Partula* on Moorea. *Evolution*, 22, 103–117.
- Nei, M., & Tajima, F. (1981). Genetic drift and estimation of effective population size. *Genetics*, 98, 625–640.
- Noel, E., Chemtob, Y., Janicke, T., Sarda, V., Pelissie, B., Jarne, P., & David, P. (2016). Reduced mate availability leads to evolution of self-fertilization and purging of inbreeding depression in a hermaphrodite. *Evolution*, 70, 625–640. <https://doi.org/10.1111/evo.12886>
- Nunney, L., & Elam, D. (1994). Estimating the effective size of conserved populations. *Conservation Biology*, 8, 175–184. <https://doi.org/10.1046/j.1523-1739.1994.08010175.x>
- Pollak, E. (1983). A new method for estimating effective population sized from allele frequency changes. *Genetics*, 104, 531–548.
- Puurtinen, M., Knott, K., Suonpaa, S., Van Ooik, T., & Kaitala, V. (2004). Genetic variability and drift load in populations of an aquatic snail. *Evolution*, 58, 749–756. <https://doi.org/10.1111/j.0014-3820.2004.tb00408.x>
- Raymond, M., & Rousset, F. (1995). GENEPOP (version 1.2): Population genetics software for exact tests and ecumenicism. *Journal of Heredity*, 86, 248–249. <https://doi.org/10.1093/oxfordjournals.jhered.a111573>
- Riquet, F., Le Cam, S., Fonteneau, E., & Viard, F. (2016). Moderate genetic drift is driven by extreme recruitment events in the invasive mollusk *Crepidula fornicata*. *Heredity*, 117, 42–50. <https://doi.org/10.1038/hdy.2016.24>
- Rousset, F. (2008). Genepop'007: A complete reimplementation of the Genepop software for Windows and Linux. *Molecular Ecology Resources*, 8, 103–106. <https://doi.org/10.1111/j.1471-8286.2007.01931.x>
- Selander, R., & Kaufman, D. (1975). Genetic structure of populations of the brown snail (*Helix aspersa*) I. Microgeographic variation. *Evolution*, 29, 385–401.
- Trouve, S., Degen, L., & Goudet, J. (2005). Ecological components and evolution of selfing in the freshwater snail *Galba truncatula*. *Journal of Evolutionary Biology*, 18, 358–370.
- Ursenbacher, S., Alvarez, C., Armbruster, G., & Baur, B. (2010). High population differentiation in the rock-dwelling land snail (*Trochulus caelatus*) endemic to the Swiss Jura Mountains. *Conservation Genetics*, 11, 1265–1271. <https://doi.org/10.1007/s10592-009-9956-3>
- Van Leeuwen, C., Huig, N., Van der Velde, G., Van Alen, T., Wagemaker, C., Sherman, C., ... Figuerola, J. (2013). How did this snail get here? Multiple dispersal vectors inferred for an aquatic invasive species. *Freshwater Biology*, 58, 88–99. <https://doi.org/10.1111/fwb.12041>
- Viard, F., Justy, F., & Jarne, P. (1997). The influence of self-fertilization and population dynamics on the genetic structure of subdivided populations: A case study using microsatellite markers in the freshwater snail *Bulinus truncatus*. *Evolution*, 51, 1518–1528. <https://doi.org/10.1111/j.1558-5646.1997.tb01475.x>
- Waples, R. (1989). A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics*, 121, 379–391.
- Waples, R., & Do, C. (2008). LDNe: A program for estimating effective population size from data on linkage disequilibrium. *Molecular Ecology Resources*, 8, 753–756. <https://doi.org/10.1111/j.1755-0998.2007.02061.x>
- Weir, B., & Cockerham, C. (1984). Estimating F-statistics for the analysis of population structure. *Evolution*, 38, 1358–1370.
- Wethington, A. R., & Dillon, R. T. Jr (1993). Reproductive development in the hermaphroditic freshwater snail, *Physa*, monitored with complementing albino lines. *Proceedings of the Royal Society of London, Series B: Biological Sciences*, 252, 109–114. <https://doi.org/10.1098/rspb.1993.0053>
- Wethington, A. R., & Dillon, R. T. Jr (1996). Gender choice and gender conflict in a non-reciprocally mating simultaneous hermaphrodite, the freshwater snail, *Physa*. *Animal Behaviour*, 51, 1107–1118. <https://doi.org/10.1006/anbe.1996.0112>
- Wethington, A. R., & Dillon, R. T. Jr (1997). Selfing, outcrossing, and mixed mating in the freshwater snail *Physa heterostrophia*: Lifetime fitness and inbreeding depression. *Invertebrate Biology*, 116, 192–199. <https://doi.org/10.2307/3226896>
- Wright, S. (1931). Evolution in Mendelian populations. *Genetics*, 16, 97–159.

**How to cite this article:** Dillon Jr RT. Volatility in the effective size of a freshwater gastropod population. *Ecol Evol*. 2018;8:2746–2751. <https://doi.org/10.1002/ece3.3912>